

## Supporting Information

# **Kraft lignin solubility and its chemical modification in deep eutectic solvents**

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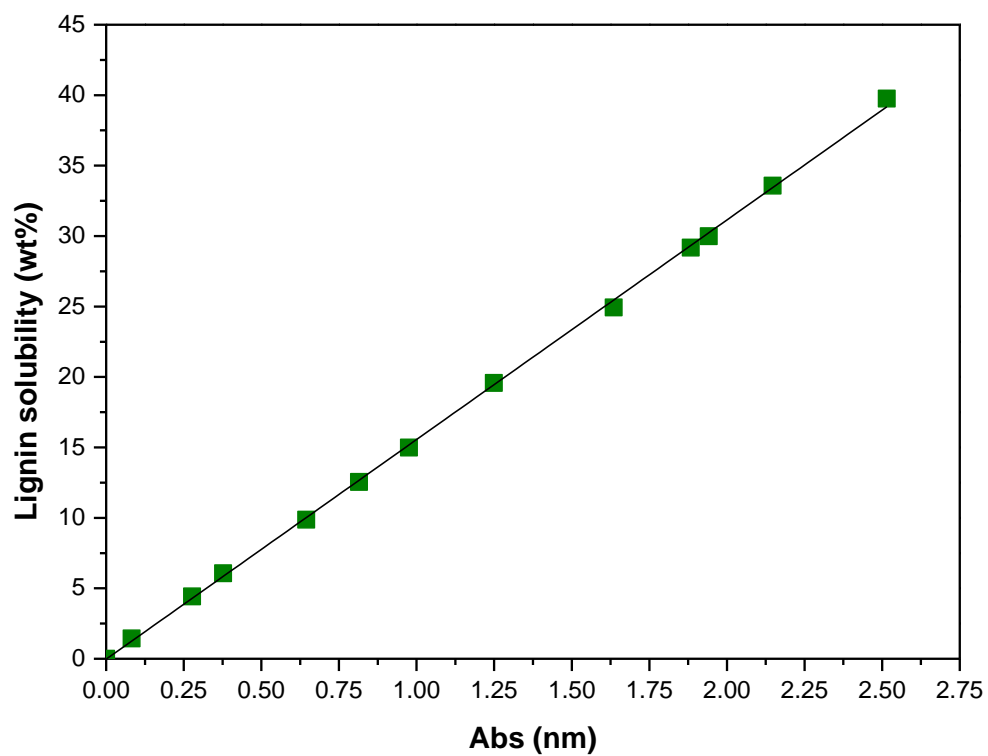
## 1. Chemicals

**Table S1.** Compounds used in this work along with their CAS Number, molecular weight (Mw), purity and supplier.

Compound	CAS Number	Mw (g mol <sup>-1</sup> )	Water content (wt %)	Purity <sup>a</sup> (wt %)	Supplier
Cholinium chloride ([Ch]Cl)	67-48-1	139.62	2.38±0.42	98.0	Acros Organics
Glycerol (GLY)	56-81-5	92.09	0.13±0.04	99.0	Acros Organics
Ethylene glycol (EGLY)	107-21-1	62.07	0.11±0.02	99.0	Fluka
1,3-propanediol (PROP)	504-63-2	76.10	0.09±0.01	98.0	Sigma
1,4-butanediol (BUT)	110-63-4	90.12	0.07±0.01	99.0	Alfa Aesar
1,5-pentanediol (PENT)	111-29-5	104.15	0.05±0.02	97.0	Alfa Aesar
1,6-hexanediol (HEXA)	629-11-8	118.16	0.08±0.01	97.0	Aldrich
L(+)-lactic acid (LacA)	79-33-4	90.08	14.8±0.05	92.0	Riedel de Haen
Oxalic acid (OxaA)	144-62-7	90.03	0.03±0.00	99.5	Merck
DL-malic acid (MaliA)	617-48-1	134.09	0.02±0.00	99.5	Panreac
DL-malonic acid (MaloA)	141-82-2	104.06	0.01±0.00	>98.0	Fluka
Maleic acid (MaleA)	110-16-7	116.07	0.01±0.00	99.0	Panreac

<sup>a</sup>reported by the supplier

## 2. LIGNIN CALIBRATION CURVE



**Figure S1.** Kraft lignin calibration curve.

### 3. KRAFT LIGNIN SOLUBILITY IN DES

**Table S2.** Solubility of Kraft lignin in DES with 5 wt % water content at 313.15 K.

DES	HBA:HBD molar ratio	Lignin solubility (wt%)
<i>Polyols</i>		
[Ch]Cl:GLY	1:2	28.56±0.89
[Ch]Cl:EGLY	1:2	32.26±0.72
[Ch]Cl:PROP	1:2	25.12±0.58
[Ch]Cl:BUT	1:2	27.61±0.56
[Ch]Cl:PENT	1:2	29.54±0.51
[Ch]Cl:HEXA	1:2	32.99±0.58
<i>Carboxylic Acids</i>		
[Ch]Cl:OxaA	1:1	23.08±0.30
[Ch]Cl:MaloA	1:1	34.10±0.04
[Ch]Cl:MaliA	1:1	24.74±0.33
[Ch]Cl:MaleA*	1:1	34.97±0.33
[Ch]Cl:OxaA	1:2	23.92±0.47
[Ch]Cl:MaloA	1:2	34.00±0.37
[Ch]Cl:MaliA	1:2	25.24±1.08
<i>References</i>		
[Ch]Cl:LacA	1:1	28.23±0.24
Water	-	0.15±0.05

\*10 wt % water content

#### 4. SHIMIZU AND MATUBAYASI HYDROTROPY MODEL

The Shimizu and Matubayasi<sup>1</sup> model was developed to describe the usual sigmoidal solubility curves found in hydrotropy and also describes information about the interactions between solute and hydrotropic molecules. The model developed by Shimizu and Matubayasi<sup>1</sup> can be described as:

$$\frac{S_L}{S_{L,0}} = \frac{1 + \left(\frac{S_L}{S_{L,0}}\right)_{max} e^{m \cdot \ln(x_H) + b}}{1 + e^{m \cdot \ln(x_H) + b}}$$

$$\text{where: } x_H = \frac{M_H}{M_H + M_{water}}$$

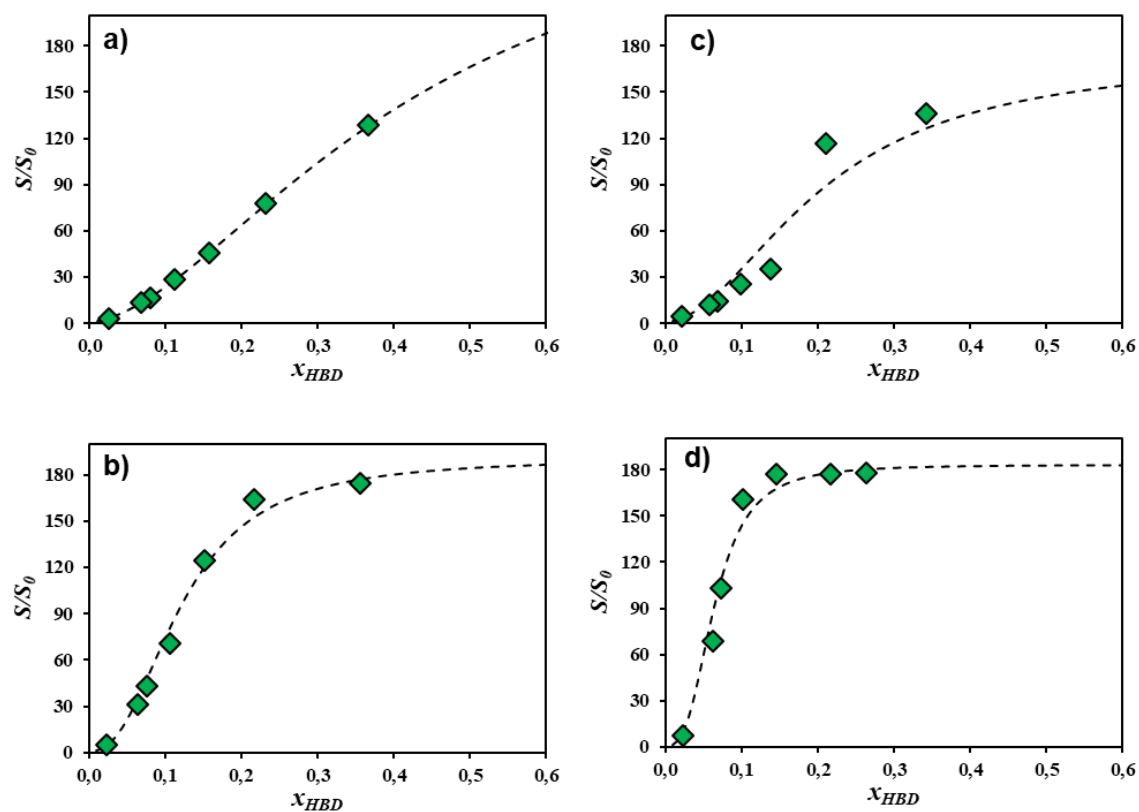
$$x_H = x_{HBD} \text{ or}$$

$$x_H = x_{ChCl} \text{ or}$$

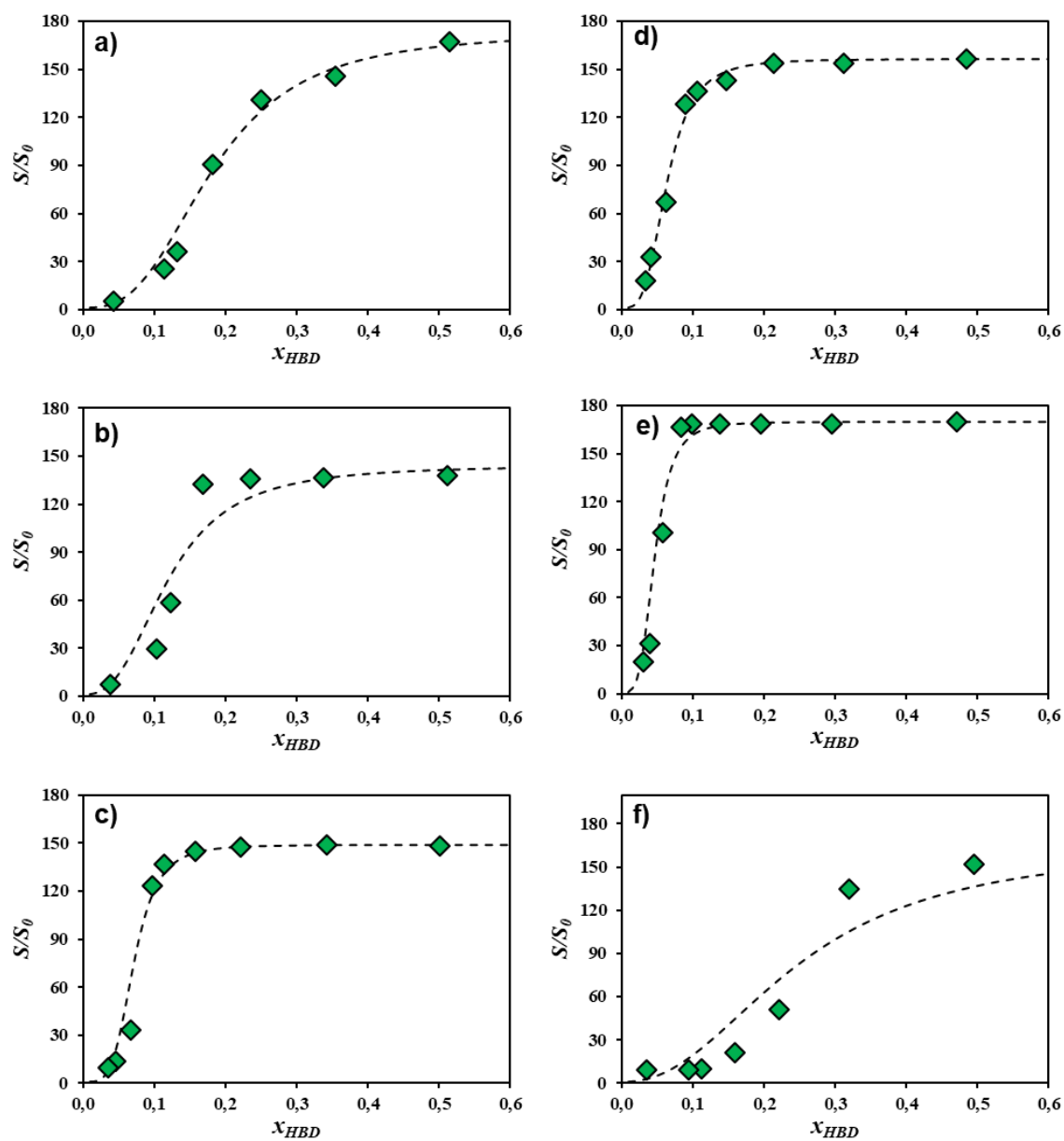
$$x_H = x_{ES} = x_{HBD} + x_{ChCl}$$

where  $S_L$  is the solubility of lignin in the system (molar fraction of the solute),  $S_{L,0}$  is the solubility of lignin in water and  $x_H$  is the molar fraction of the hydrotrope, in this case being HBD, or chlorine chloride or the sum of the two in the case of ES. Note that  $x_H$  is the molar fraction of the hydrotrope on a solute free base. The term  $S_L/S_{L,0}$  represents the relative solubility of lignin in the solution about solubility in pure water. The parameters  $m$  and  $b$  represent the molecular interactions between solute and hydrotrope, more specifically,  $m$  represents the number of hydrotrope molecules in the vicinity of the solute.

After determining the parameters  $m$  and  $b$ , it is possible to outline the model curve adjusted to the experimental data.



**Figure S2.** Solubility enhancement of lignin ( $S_L/S_{L,0}$ ) in DES aqueous solutions with the following HBDs: a) OxaA, b) MaloA, c) MaliA, and d) MaleA fitted using the Shimizu and Matubayasi hydrotrophy model (---).<sup>1</sup>



**Figure S3.** Solubility enhancement of lignin ( $S_L/S_{L,0}$ ) in DES aqueous solutions with the following HBDs: a) ethylene glycol, b) 1,3-propanediol, c) 1,4-butanediol, d) 1,5-pentanediol and e) 1,6-hexanediol and f) glycerol fitted using the Shimizu and Matubayasi hydrotrophy model (- -).<sup>1</sup>

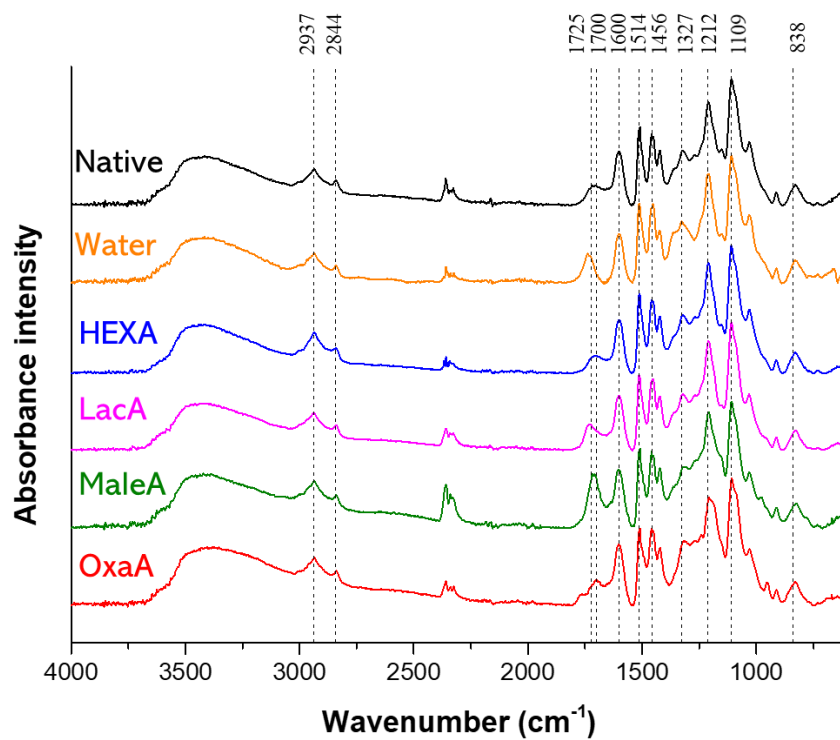


## 5. LIGNIN CHARACTERIZATION

### 5.1. FTIR-ATR

**Table S3.** FTIR vibrational bands/regions and corresponding assignments for Kraft lignin.

Vibrational band/region (cm <sup>-1</sup> )	Assignments
3200-3500	O-H vibrations <sup>2</sup>
2844-2980	Aliphatic C-H and CH <sub>2</sub> stretching vibrations <sup>2</sup>
1775-1750	C=O of esters, ketones, aldehydes and acids. ( C=O stretching, non conjugated <sup>3</sup>
1731	C=O stretching in xylan, C=O stretching of acetyl or carboxylic acid <sup>4,5</sup>
1700	Unconjugated C=O (ketone, carboxyl or ester stretching) <sup>4,6</sup>
1600-1690	Aromatic skeletal vibration <sup>7-9</sup>
1514	Aromatic skeletal vibration <sup>7-9</sup>
1456	Aromatic skeletal vibration and C-H deformations <sup>7-9</sup>
1425	Aromatic skeletal vibrations combined with C-H in-plane deformation <sup>7-9</sup>
1370-1365	C-H deformation (aliphatic C-H Stretch in CH <sub>3</sub> , not in OMe; phen. OH <sup>10</sup>
1327-1365	Syringyl unit breathing with C=O stretching and condensed Guaiacyl rings <sup>7-9</sup>
1241	Guaiacyl ring breathing C-O stretch in lignin and for C-O linkage in guaiacyl aromatic methoxyl groups <sup>11</sup>
1212	C-C plus C-O plus C=O stretch; Guaiacyl condensed > Guaiacyl etherified <sup>7-9</sup>
1152	C-O-C vibration (Cellulose and hemicellulose) <sup>11</sup>
1109	Contribution of C-H in a plane deformation, C=O stretching of syringyl units and secondary alcohols <sup>7-9</sup>
1040	Aromatic C H in-plane deformation (Guaiacyl > Syringyl) plus C-O deformation in primary alcohols plus C=O stretch (unconjugated) <sup>7-9</sup>
979	C-O valance vibration; aromatic C-H in plane deformation <sup>12</sup>
925	Aromatic C-H out-of-plane <sup>7</sup>
838	Aromatic C-H out-of-plane deformation in Guaiacyl and Syringyl units <sup>13</sup>



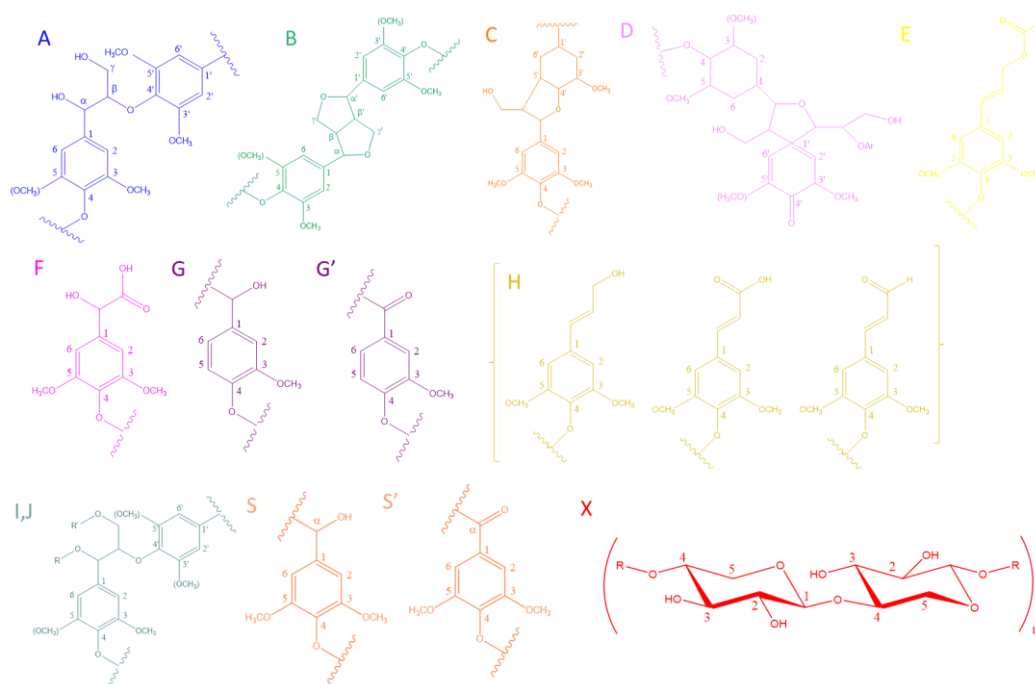
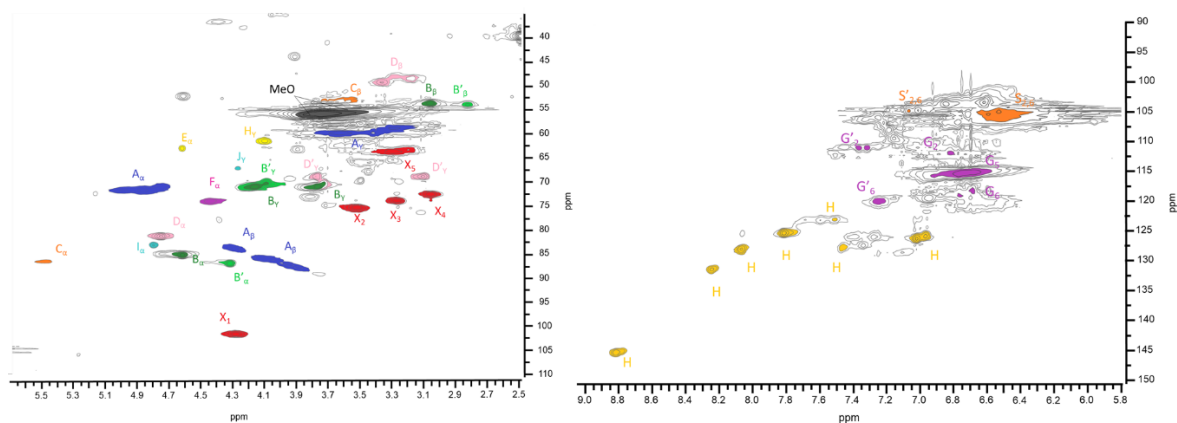
**Figure S4.** FTIR-ATR spectra of Kraft lignin and recovered lignins from treatments with [Ch]Cl:OxaA, [Ch]Cl:LacA, [Ch]Cl:Hexa, [Ch]Cl:MaleA and water at 393.15 K for 6 h.

## 5.2. 2D HSQC NMR

The structural analysis of lignin samples was complemented with the 2D HSQC NMR technique to better understand the impact of thermal treatments with DES. The HSQC oxygenated and aromatic regions of the Kraft lignin along with the representation of identified lignin subunits are shown in Figure S5 to suitably address its chemical characterization. The  $^{13}\text{C}$ - $^1\text{H}$  cross signal assignments are described in Table S4 according to data reported in literature.<sup>14-20</sup>

In the oxygenated aliphatic region ( $\delta_{\text{C}}\text{-}\delta_{\text{H}}$  35-110/2.5-5.7 ppm), the major lignin subunits, such as alkyl-aryl ether structures ( $\beta$ -O-4), resinols ( $\beta$ - $\beta$ ), phenylcoumaran ( $\beta$ -1), spirodienone ( $\beta$ -5), were identified in Kraft Lignin, while an intense signal at 56.1/3.75 ppm represents the C-H cross assignment of methoxy groups in lignin. On the other hand, cross signals associated with xylan backbone (X) were also found representing a contamination of residual xylan in Kraft lignin, which is a common trait in this technical lignin.<sup>15</sup>

In the aromatic region ( $\delta_{\text{C}}\text{-}\delta_{\text{H}}$  90-160/5.8-9.0 ppm), the cross signals at 104.88/7.07 and 105.22/6.47 ppm refer to the  $\text{C}_{2,6}\text{-H}_{2,6}$  correlations in non-oxidized ( $\text{S}_{2,6}$ ) and oxidized ( $\text{S}'_{2,6}$ ) syringyl type units, respectively. Similar assignments can be made to guaiacyl type units, including non-oxidized at 111.00/6.95 ppm ( $\text{G}_2$ ), 115.00/6.75 ppm ( $\text{G}_5$ ), 119.00/6.75 ppm ( $\text{G}_6$ ); and oxidized at 111.04/7.33 ( $\text{G}'_2$ ) and 120.14/7.24 ( $\text{G}'_6$ ) ppm.<sup>21</sup> The signal range between 122.00-130.00/7.0-8.0 ppm should correspond to  $\alpha$ -carbonyl/carboxyl position in syringyl and guaiacyl units<sup>14</sup> as well as to cinnamyl structures with alcohols, aldehydes, and acids as end groups (H).

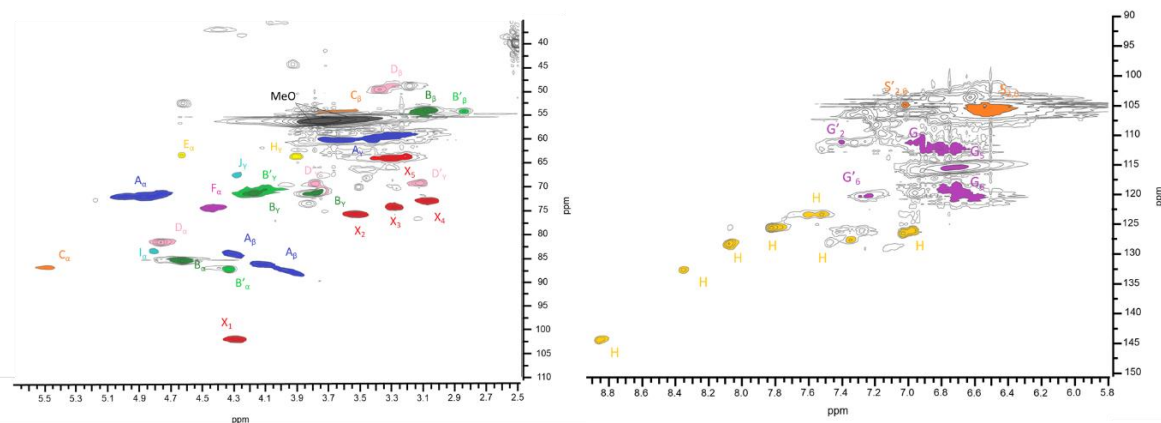


**Figure S5.** 2D HSQC NMR of Kraft lignin with corresponding  $^{13}\text{C}$ - $^1\text{H}$  cross signal assignments to main lignin subunits: (A)  $\beta$ -O-4' linkages; (B) Resinol ( $\beta$ - $\beta'$ ); (C) Phenylcoumaran ( $\beta$ -5'); (D) Spirodienone ( $\beta$ -1'); (E) cinnamyl acetate end-groups; (F) Ar-CHOH-COOH unit (C $\alpha$ -H $\alpha$ ); (G) Guaiacyl unit; (G') oxidized Guaiacyl unit with a C $\alpha$  ketone or carboxyl group; (H) p-hydroxycinnamyl structures (alcohol, aldehyde or carboxylic end groups); (I)  $\beta$ -O-4' substructure C $\alpha$  etherified to carbohydrate (R, polysaccharide; R', H); (J)  $\beta$ -O-4' substructure C $\gamma$  etherified to carbohydrate (R, H; R', polysaccharide); (S, S') Syringyl unit; (S', S'') oxidized syringyl unit with a C $\alpha$  ketone (phenolic); (X) Xylan.

**Table S4.** Assignments of Kraft lignin C-H correlation signals in the HSQC spectra.

Labels	$\delta C$	$\delta H$	Assignment
D $\beta$	49.61	3.39	C $\beta$ -H $\beta$ in $\beta$ -1' spirodienone substructures (D)
B $\beta$	53.50	3.07	C $\beta$ -H $\beta$ in resinol substructures (B)
C $\beta$	53.71	3.46	C $\beta$ -H $\beta$ in $\beta$ -5 phenylcoumaran
B' $\beta$	54.20	2.82	C $\beta$ -H $\beta$ in epiresinol substructures (B)
MeO	56.06	3.75	C-H in methoxyls
A $\gamma$	59.00	3.2	C $\gamma$ -H $\gamma$ in $\beta$ -O-4' substructures (A)
A $\gamma$	59.87	3.71	C $\gamma$ -H $\gamma$ in $\beta$ -O-4' substructures (A)
H $\gamma$	61.59	4.11	C $\gamma$ -H $\gamma$ in p-hydroxycinnamyl alcohol
E $\alpha$	63.16	4.64	C $\alpha$ -H $\alpha$ in p-hydroxycinnamyl alcohol
C $\gamma$	63.26	3.89	C $\gamma$ -H $\gamma$ in $\beta$ -5 phenylcoumaran
X5	63.63	3.28	C5-H5 in xylan
J $\gamma$	66.88	4.28	C $\gamma$ -H $\gamma$ in $\beta$ -O-4' C $\alpha$ - etherified with carbohydrate
D $\gamma'$	68.89	3.79	C $\gamma$ -H $\gamma$ in $\beta$ -1' spirodienone substructures (D)
D $\gamma'$	68.92	3.13	C $\gamma$ -H $\gamma$ in $\beta$ -1' spirodienone substructures (D)
B' $\beta$	70.56	4.09	C $\beta$ -H $\beta$ in epiresinol substructures (B)
B $\gamma$	71.07	3.79	C $\gamma$ -H $\gamma$ in resinol substructures (B)
B $\gamma$	71.14	4.2	C $\gamma$ -H $\gamma$ in resinol substructures (B)
A $\alpha$	71.62	4.91	C $\alpha$ -H $\alpha$ in $\beta$ -O-4' substructures (A)
X4	72.85	3.06	C4-H4 in xylan
F $\alpha$	74.18	4.43	C $\alpha$ -H $\alpha$ in Ar-CHOH-COOH unit
X3	74.28	3.28	C3-H3 in xylan
X2	75.84	3.53	C2-H2 in xylan
B' $\alpha$	81.36	4.78	C $\alpha$ -H $\alpha$ in epiresinol substructures (B)
C $\alpha$	82.92	5.51	C $\alpha$ -H $\alpha$ in $\beta$ -5 phenylcoumaran
I $\alpha$	83.41	4.83	C $\alpha$ -H $\alpha$ in $\beta$ -O-4' C $\alpha$ - etherified with carbohydrate
A $\beta$	83.96	4.31	(G) C $\beta$ H $\beta$ in $\beta$ -O-4' linked to G units
B $\alpha$	85.3	4.67	C $\alpha$ -H $\alpha$ in resinol substructures (B)
D $\beta'$	86.89	4.44	C $\beta$ -H $\beta$ in $\beta$ -1' spirodienone substructures (D)
B' $\alpha$	87.22	4.33	C $\alpha$ -H $\alpha$ in epiresinol substructures (B)
A $\beta$	87.23	3.69	(S) C $\beta$ -H $\beta$ in $\beta$ -O-4' linked to S units
X1	101.8	4.29	C1-H1 in xylan
S'2.6	104.88	7.07	C2.6 - H2.6 in oxidized Syringyl units (S')

S2.6	105.22	6.47	C <sub>2,6</sub> - H <sub>2,6</sub> in Syringyl units (S)
G2	111.00	6.95	C <sub>2</sub> -H <sub>2</sub> in Guaiacyl units (G)
G2'	111.04	7.33	C <sub>2</sub> -H <sub>2</sub> in oxidized Guaiacyl units (G)
G5	115.00	6.75	C <sub>5</sub> -H <sub>5</sub> in Guaiacyl units (G)
G6	119.00	6.75	C <sub>6</sub> -H <sub>6</sub> in Guaiacyl units (G)
G6'	120.14	7.24	C <sub>6</sub> -H <sub>6</sub> in oxidized Guaiacyl units (G)
H	122.98	7.56	Cinnamyl alcohols, aldehydes or acids (end groups)
H	125.53	7.79	Cinnamyl alcohols, aldehydes or acids (end groups)
H	126.16	4.00	Cinnamyl alcohols, aldehydes or acids (end groups)
H	127.01	7.35	Cinnamyl alcohols, aldehydes or acids (end groups)
H	128.42	8.09	Cinnamyl alcohols, aldehydes or acids (end groups)
H	131.29	8.23	Cinnamyl alcohols, aldehydes or acids (end groups)
H	145.73	8.8	Cinnamyl alcohols, aldehydes or acids (end groups)



**Figure S6.** 2D HSQC NMR of recovered lignins from thermal treatment with water at 393.15 K for 6 h.

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